

Patient Instructions for Stool Collection and Handling

The following instructions are intended to help you collect the correct specimen for the test your physician has requested. Read all the instructions carefully, make sure you are prepared, and follow each step described below to ensure proper collection.

You have been provided the following stool collection containers:

- For Ova and Parasite- Zinc-PVA/Formalin* (plastic vials with pink and blue caps)
 - For Fecal Fat Qualitative Testing, C. Difficile testing, and WBC smear (sterile, collection cup)
 - For GI PCR and stool culture- C&S Medium (plastic vial with yellow cap)
 - For IFOB, Fecal Occult Blood test - (plastic sampling bottle with green cap)
1. Antacids, barium bismuth, anti-diarrhea medication or oily laxatives should not be used prior to collection of the specimen.
 2. Collect the number of samples ordered by the physician. Do not collect more than one sample per day if provided multiple containers of the same color.
 3. The stool sample should not be contaminated with urine or toilet paper.
 4. Collect stool in a clean, dry wide mouthed container. A clean, dry piece of newspaper or wax paper may also be used.
 5. Once stool is obtained, use a tongue depressor or spoon built into the vial cap to remove a portion of the stool and place into the container provided. Collect any mucus or blood with the specimen(s).
 - a. Yellow, Blue and Pink capped container add stool to the "Red Fill Line".
 - b. Sterile plastic container transfer portions of the stool but do not overfill.
 6. For IFOB, open the sampling bottle cap by turning to the left and pulling upwards. Scrape the surface of the fecal sample with the sample probe, stabbing the stool 5-6 different times. Cover the grooved portion of the sample probe completely with stool. If loose stool, sample should be collected the same way by completely covering the grooved portion of the probe. Insert sample probe into sampling bottle and screw cap tightly.
 7. Tighten the lid on all containers to prevent leakage. Shake the yellow, pink and blue container to mix the contents.
 8. Complete the information requested on the container label. Make sure you include your full name, and the date and time you collected the specimen.
 9. Place container(s) in plastic biohazard bag and seal.
 10. Wash your hands thoroughly after collecting the specimen.
 11. Specimen should be transported to the lab as soon as possible. If a delay of more than two hours is expected, refrigerate the sample.

***Caution: 10% FIXATIVE CONTAINS FORMALIN**

10% Formalin is a poison. Toxic by inhalation and if swallowed. Irritating to the eyes, respiratory system, and skin. May cause sensitization by inhalation or by skin contact. Risk of serious damage to eyes.